



---

Howe, RA, Monk, A, Wootton, M, Walsh, TR and Enright, Mark (2004) Vancomycin Susceptibility within Methicillin-resistant *Staphylococcus aureus* Lineages. *Emerging Infectious Diseases*, 10 (5). pp. 855-857. ISSN 1080-6040

---

**Downloaded from:** <https://e-space.mmu.ac.uk/621205/>

**Version:** Published Version

**Publisher:** Centers for Disease Control and Prevention

**DOI:** <https://doi.org/10.3201/eid1005.030556>

**Usage rights:** Creative Commons: Attribution 3.0

Please cite the published version

<https://e-space.mmu.ac.uk>

---

# Vancomycin Susceptibility within Methicillin-resistant *Staphylococcus aureus* Lineages

Robin A. Howe,\* Alastair Monk,† Mandy Wootton,\* Timothy R. Walsh,‡ and Mark C. Enright†

Methicillin-resistant *Staphylococcus aureus* (MRSA) with reduced vancomycin susceptibility vancomycin-intermediate *S. aureus* (VISA) has been reported from many countries. Whether resistance is evolving regularly in different genetic backgrounds or in a single clone with a genetic predisposition, as early results suggest, is unclear. We have studied 101 MRSA with reduced vancomycin susceptibility from nine countries by multilocus sequence typing (MLST), characterization of *SCCmec* (staphylococcal chromosomal cassette *mec*), and *agr* (accessory gene regulator). We found nine genotypes by MLST, with isolates within all five major hospital MRSA lineages. Most isolates (88/101) belonged to two of the earliest MRSA clones that have global prevalence. Our results show that reduced susceptibility to vancomycin has emerged in many successful epidemic lineages with no clear clonal disposition. Increasing antimicrobial resistance in genetically distinct pandemic clones may lead to MRSA infections that will become increasingly difficult to treat.

Methicillin-resistant *Staphylococcus aureus* (MRSA) is a major problem around the world, causing hospital-acquired infections and, more recently, infections in the community (1,2). The glycopeptides, particularly vancomycin, have been the mainstays of therapy for MRSA, and the emergence of resistance to these agents is of great concern.

The first *S. aureus* with reduced vancomycin susceptibility (vancomycin MIC  $\geq 8$   $\mu\text{g/mL}$ ) was isolated in 1997 (3,4), and similar isolates have since been discovered in several countries. These vancomycin-intermediate *S. aureus* (VISA) isolates are relatively rare; a recent review found 21 VISA described in the literature (5). However, strains of *S. aureus* have been described that are vancomycin-susceptible by conventional testing but have a subpopulation of resistant cells. These heterogeneous

VISA (hVISA) are more common; reports from around the world indicate that 0.5%–20% of MRSA are heteroresistant (5). The clinical importance of hVISA is debatable, but evidence shows that they are precursors of VISA, and they have been implicated in treatment failure in deep-seated infections (6,7).

A study of early VISA strains that used multilocus sequence typing (MLST) and analysis of the *SCCmec* region suggested that they were all descended from the New York/Japanese (8,9) pandemic MRSA clone (10); the first high-level vancomycin-resistant isolates that have acquired the *vanA* gene cassette from enterococci are also members of this clone (F. Tenover, pers. comm.). Researchers have suggested that isolates of the New York/Japanese pandemic MRSA clone may be predisposed to become vancomycin resistant, perhaps because of loss-of-function mutations in the *agr* (accessory gene regulator) gene (11). We analyzed the genetic backgrounds of a geographically diverse sample of VISA and hVISA to investigate the evolutionary history of such strains.

## Materials and Methods

We collected 101 isolates of MRSA with reported heterogeneous or homogeneous resistance to vancomycin (MIC  $\geq 8$  mg/L) from China ( $n = 1$ ), France (31), Japan (2), Norway (14), Poland (13), Sweden (1), United Kingdom (28), and the United States (11). Antimicrobial susceptibility tests were performed by the agar dilution method of the National Committee for Clinical Laboratory Standards. Isolates were described as VISA if they fulfilled the three criteria adopted by the Centers for Disease Control and Prevention, that is, broth microdilution vancomycin MIC of 8 to 16 mg/L, MIC  $\geq 6$  mg/L on E-test, and growth on brain-heart infusion agar containing 6 mg/L vancomycin (12). Isolates with heterogeneous resistance to vancomycin were confirmed by using population analysis profiling followed by measuring the area under the curve (PAP-AUC), as described previously (13). The prototypic

---

\*Southmead Hospital, Bristol, United Kingdom; †University of Bath, Bath, United Kingdom; and ‡University of Bristol, Bristol, United Kingdom

hVISA strain MU3 was used as a standard, and isolates with an AUC  $\geq 0.9$  compared to MU3 were described as hVISA.

MLST was performed as described previously (10). The seven housekeeping gene sequences were compared to known alleles in the MLST database (available from <http://www.mlst.net>), and the resulting allelic profiles (which define sequence types, STs) were used to interrogate the databases for matches within records of the 988 isolates held there. The MLST databases contain molecular and epidemiologic data on *S. aureus* isolates from carriage and disease, including examples of all major MRSA clones (10). Data from this study were added to the *S. aureus* MLST database, and the entire dataset was analyzed by using the BURST algorithm to assign isolates to clonal complexes (CCs), which are lineages containing genetically related isolates (sharing 100% genetic identity at  $\geq 5/7$  loci used). Polymerase chain reaction (PCR) analysis of the *ccr* (chromosomal cassette recombinase) and *mec* (methicillin resistance) regions was performed to discriminate the four main SCCmec types (I–IV) on the basis of combinations of the two regions. Conventional PCR was used to detect SCCmec I–III by using the primers described in Ito et al. (14) and SCCmec IV by using those described by Daum et al. (15). These results were confirmed using the multiplex method of Oliveira et al. (16). Detection of *agr* subgroups

I–IV was performed by PCR of the region surrounding *agrD*, which codes for an autoinducing peptide, according to the method of Peacock et al. (17).

## Results and Discussion

The results are shown in the Table. PAP-AUC values for the isolates varied from 0.9 to 3.01 and 91/101 isolates were designated hVISA on the basis of a PAP-AUC value  $\geq 0.9$ . Nine isolates were designated as VISA.

From the genotyping results, strains were divided into clonal complexes, which can be subdivided according to sequence type (ST) and SCCmec differences. The clonal complexes CC5, CC8, CC22, CC30, and CC45 represent the five pandemic MRSA lineages that have been previously described (10). Our results show that hVISA has arisen in all five of these pandemic clones and that VISA has so far developed in CC5 and CC8. The three most common MRSA clones present in the United Kingdom (EMRSA-3, EMRSA-15, EMRSA-16) (18) are included within these lineages, and reduced vancomycin susceptibility has been identified in all of these clones. All lineages displayed resistance to multiple antimicrobial classes, and only the new oxazolidinone linezolid was active against all strains.

Only *agr* subgroups (alleles) I and II were found in isolates in this study with 7/9 VISA and 57/92 hVISA having

Table. Characteristics of study isolates with reduced vancomycin susceptibility<sup>a</sup>

Genotype					Vancomycin resistance phenotype	PAP-AUC	Country of origin	Antimicrobial susceptibility <sup>*†</sup>				
CC	ST	SCCmec	Clonal type	<i>agr</i> type	(no. of strains)			Lzd	Syn	Gen	Cip	Rif
5	5	I	EMRSA-3	II	hVISA (1)	0.98	UK	S	S	R	R	R
				I	VISA (1)	1.9	USA	S	S	R	R	S
5	5	II	New York/Japanese	II	hVISA (10)	0.97–1.23	Japan, Sweden, France, Poland, UK, USA, Norway	S	S	S/R	S/R	S/R
				I or II	VISA (3)	1.4–1.92	USA	S	S	S	R	S/R
5	5	IV	Pediatric	I or II	hVISA (3)	1.19–1.32	UK	S	S	S	S/R	S/R
5	5	NT		I	VISA (1)	1.44	France	S	S	R	R	R
8	8	I		II	hVISA (3)	0.92–1.32	France, UK, Norway	S	S	R	R	R
8	8	II	Irish-1	II	hVISA (3)	1.04–1.2	France, USA, Norway	S	S	S/R	R	S/R
8	8	IV	EMRSA-4, -6	I	hVISA (11)	0.94–1.24	France, USA	S	S/R	S/R	R	S/R
22	22	IV	EMRSA-15	I or II	hVISA (7)	0.9–1.25	UK	S	S	S	R	S/R
25	25	NT		I	hVISA (1)	1.13	UK	S	S	R	R	R
30	36	II	EMRSA-16	II	hVISA (3)	0.92–1.17	UK	S	S	R	R	R
45	45	II		I	hVISA (1)	1	USA	S	S	R	R	S
8	239	I or II	Brazilian/Portuguese	I or II	hVISA (10)	0.9–1.22	France, Poland, China, Norway, UK	S	S	R	S/R	S/R
				I	VISA (3)	1.44–3.01	France, Poland, UK	S	S/R	R	R	S/R
8	239	NT		I	hVISA (1)	0.92	France	S	S	R	R	R
8	246	NT		I	hVISA (1)	1.13	Norway	S	S	R	R	R
8	247	I	Iberian	I	VISA (1)	1.57	UK	S	S	R	R	R
				I or II	hVISA (37)	0.9–1.33	France, Poland, UK, Norway	S	S	R	R	S/R

<sup>a</sup>S, susceptible; R, resistant; NT, nontypeable; PAP-AUC, population analysis profiling followed by measuring the area under the curve; Lzd, linezolid; Syn, synergicid; Gen, gentamicin; Cip, ciprofloxacin; Rif, rifampin; CC, clonal complex; ST, sequence type; EMRSA, methicillin-resistant *Staphylococcus aureus* found in the United Kingdom (UK); hVISA, heterogeneous vancomycin-intermediate *S. aureus*; USA, United States of America.

*agr* I. Within the 14 clones in this study, the proportion of isolates with particular *agr* alleles was variable. The presence of both *agr* I and *agr* II among VISA/hVISA, even in genetically similar isolates, suggests that the genes for the *agr* system are horizontally transferred. Sakoulas et al. reported an association of *agr* II with the development of vancomycin resistance (11). Our results show that VISA/hVISA also emerged in strains with *agr* I.

Molecular analyses of VISA isolates to date have focused on isolates from the United States and Japan, and results have indicated that all strains belong to the New York/Japanese MRSA clone. In our study, we found that hVISA isolates have emerged from every lineage that has produced pandemic MRSA clones, and VISA isolates have emerged in two of five lineages, in all likelihood from hVISA precursor isolates.

Increasing drug resistance in clones that are multidrug resistant and adapted to spread and cause serious disease can do much damage in the modern hospital environment. We have shown that reduced vancomycin susceptibility has emerged in genetically and phenotypically diverse MRSA clones throughout the world. This finding suggests that vancomycin resistance has the potential to become a widespread problem in MRSA strains already resistant to multiple antimicrobial agents.

This work was funded by the Wellcome Trust. M.C.E. is a Royal Society University Research Fellow.

Dr. Howe is a consultant microbiologist at Southmead Hospital, Bristol, and Clinical Lecturer at Bristol University. His research interests include many areas of clinical microbiology, particularly the mechanisms of antimicrobial resistance in bacterial pathogens.

## References

- Chambers HF. The changing epidemiology of *Staphylococcus aureus*? *Emerg Infect Dis* 2001;7:178–82.
- Herold BC, Immergluck LC, Maranan MC, Lauderdale DS, Gaskin RE, Boyle-Vavra S, et al. Community-acquired methicillin-resistant *Staphylococcus aureus* in children with no identified predisposing risk. *JAMA* 1998;279:593–8.
- Centers for Disease Control and Prevention. Update: *Staphylococcus aureus* with reduced susceptibility to vancomycin—United States, 1997. *MMWR Morb Mortal Wkly Rep* 1997;46:813–5.
- Hiramatsu K, Hanaki H, Ino T, Yabuta K, Oguri T, Tenover FC. Methicillin-resistant *Staphylococcus aureus* clinical strain with reduced vancomycin susceptibility. *J Antimicrob Chemother* 1997;40:135–6.
- Walsh TR, How RA. The prevalence and mechanisms of vancomycin resistance in *Staphylococcus aureus*. *Annu Rev Microbiol* 2002;56:657–75.
- Ariza J, Pujol M, Cabo J, Pena C, Fernandez N, Linares J, et al. Vancomycin in surgical infections due to methicillin-resistant *Staphylococcus aureus* with heterogeneous resistance to vancomycin. *Lancet* 1999;353:1587–8.
- Moore MR, Perdreau-Remington F, Chambers HF. Vancomycin treatment failure associated with heterogeneous vancomycin-intermediate *Staphylococcus aureus* in a patient with endocarditis and in the rabbit model of endocarditis. *Antimicrob Agents Chemother* 2003;47:1262–6.
- Ito T, Katayama Y, Hiramatsu K. Cloning and nucleotide sequence determination of the entire *mec* DNA of pre-methicillin-resistant *Staphylococcus aureus* N315. *Antimicrob Agents Chemother* 1999;43:1449–58.
- Oliveira DC, Tomasz A, de Lencastre H. The evolution of pandemic clones of methicillin-resistant *Staphylococcus aureus*: identification of two ancestral genetic backgrounds and the associated *mec* elements. *Microb Drug Res* 2001;7:349–61.
- Enright MC, Robinson DA, Randle G, Feil EJ, Grundmann H, Spratt BG. The evolutionary history of methicillin-resistant *Staphylococcus aureus* (MRSA). *Proc Natl Acad Sci U S A* 2002;99:7687–92.
- Sakoulas G, Eliopoulos GM, Moellering RC Jr, Wannersten C, Venkataraman L, Novick RP, et al. Accessory gene regulator (*agr*) locus in geographically diverse *Staphylococcus aureus* isolates with reduced susceptibility to vancomycin. *Antimicrob Agents Chemother* 2002;46:1492–502.
- Tenover FC, Biddle JW, Lancaster MV. Increasing resistance to vancomycin and other glycopeptides in *Staphylococcus aureus*. *Emerg Infect Dis* 2001;7:327–32.
- Wootton M, Howe RA, Hillman R, Walsh TR, Bennett PM, MacGowan AP. A modified population analysis profile (PAP) method to detect hetero-resistance to vancomycin in *Staphylococcus aureus* in a UK hospital. *J Antimicrob Chemother* 2001;47:399–403.
- Ito T, Katayama Y, Asada K, Mori N, Tsutsumimoto K, Tiensasitorn C, et al. Structural comparison of three types of staphylococcal cassette chromosome *mec* integrated in the chromosome in methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother* 2001;45:1323–36.
- Daum RS, Ito T, Hiramatsu K, Hussain F, Mongkolrattanothai K, Jamklang M, et al. A novel methicillin-resistance cassette in community-acquired methicillin-resistant *Staphylococcus aureus* isolates of diverse genetic backgrounds. *J Infect Dis* 2002;186:1344–7.
- Oliveira DC, de Lencastre H. Multiplex PCR strategy for rapid identification of structural types and variants of the *mec* element in methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother* 2002;46:2155–61.
- Peacock SJ, Moore CE, Justice A, Kantzanou M, Story L, Mackie K, et al. Virulent combinations of adhesin and toxin genes in natural populations of *Staphylococcus aureus*. *Infect Immun* 2002;70:4987–96.
- Epidemic methicillin resistant *Staphylococcus aureus*. *Commun Dis Rep CDR Wkly* 1997;7:191.

Address for correspondence: Mark C. Enright, Department of Biology and Biochemistry, University of Bath, Bath, BA2 7AY, UK; fax: +44-1225-386779; email: m.c.enright@bath.ac.uk

Use of trade names is for identification only and does not imply endorsement by the Public Health Service or by the U.S. Department of Health and Human Services.

All material published in *Emerging Infectious Diseases* is in the public domain and may be used and reprinted without special permission; proper citation, however, is appreciated.